

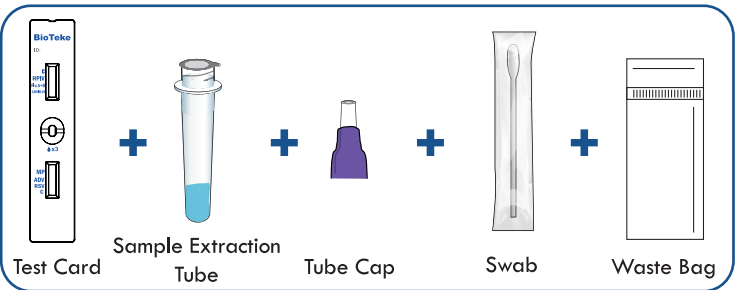
SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit

BioTeke
USER INSTRUCTION



1. Read this instruction guide carefully.
2. Have ready a watch (or a clock/timer), tissues and either hand sanitizer or soap and warm water.
3. Check the test kit contents to make sure that nothing is damaged or broken.

-For anterior nasal swabs or oropharyngeal swabs.
-Please read the instructions carefully before you begin testing.



Note: Test cards kept at low temperature should be restored to room temperature before opening to avoid moisture absorption.

Note: Materials required but not provided.

- (1) Watch (or a clock/timer),
- (2) Tissues,
- (3) Hand sanitizer / soap.

1

Wash your hands thoroughly for at least 20 seconds before the test.



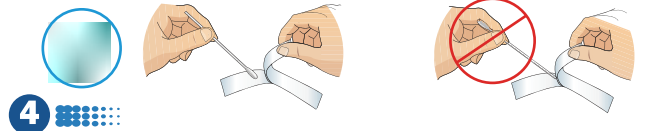
2

Put the tube into the kit box holder and gently peel off the aluminum foil seal.

3

Either of the anterior nasal swab collection and the oropharyngeal swab collection can be chosen. Once the collection is complete, the later test steps are the same.

Remove the swab from its wrapper and take out the swab by holding the handle. Do not touch the fabric tip of the swab with your hands.

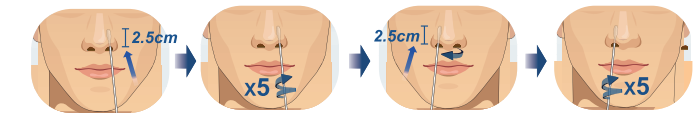


4

Anterior nasal swab collection:

NOTE: Please blow your nose before swabbing for specimen collection.

Gently insert the swab for less than one inch (about 2.5cm) into one nostril. Slowly rub the swab against all of the inside of your nose. Make at least 5 big circles. Do not just spin the swab. Repeat this step in your other nostril using the same swab.



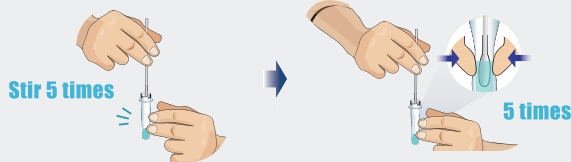
NOTE: With children, the maximum depth of insertion into the nostril may be less than 3/4 inch, please adjust according to age.

Oropharyngeal swabs collection:

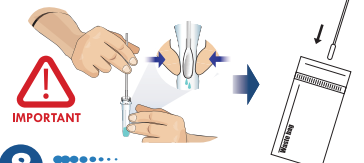
Oropharyngeal swab collection: Insert the swab in the mouth completely into the pharynx, centering on the red swelling of the pharynx wall and upper anterior tonsils. Wipe both sides of pharyngeal tonsils and pharynx posterior wall with moderate force, avoid touching the tongue, and remove the swab.



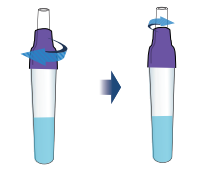
5 Insert the swab into the sample tube. Touch the bottom of the sample tube with the swab tip, and stir at least 5 times. Squeeze the swab in the tube through the outer wall of the tube by fingers 5 times.



6 Remove the swab by rotating against the sample tube while squeezing the sides of the tube to release the liquid from the swab. Remove and discard the swab.

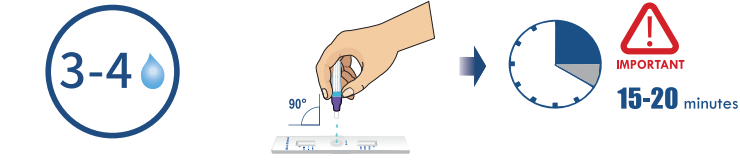


7 Screw the purple tube cap onto the sample tube and then unscrew the top white cap.



8

Open the pouch and take out the Test Card. Place it on a flat, dry and clean surface. Turn the tube integrated dropper cap upside down and slowly squeeze 3 or 4 drops into the sample well of the Test Card.



9 Results Interpretation

NOTE: The test results should not be read after 30 minutes.



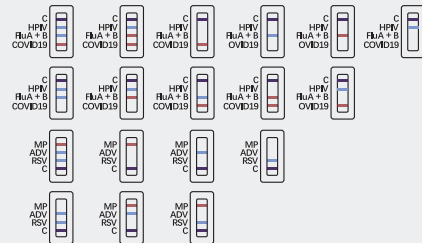
(Positive)

Two colored (C) lines appear in two separate windows, If the other line appears, the corresponding pathogen is positive.

Special Flu A+B with red color representing influenza A virus and blue color representing influenza B virus.

Note: A positive result means that you are likely to be infected with SARS-CoV-2/Influenza A virus/ Influenza B virus/Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Mycoplasma pneumoniae.

Note: Test results should always be interpreted in the context of clinical observations and epidemiological data when making final diagnoses and patient management decisions.



(Negative)

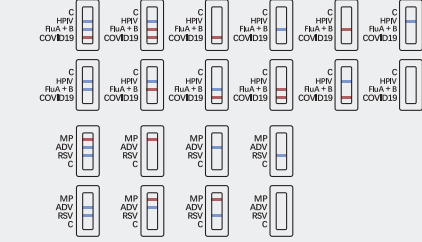
Two colored (C) lines appear in each window, and if no other lines appear, the corresponding pathogen is negative. However, a negative result does not exclude the absence of SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/ Mycoplasma pneumoniae infection and should not be used as the sole basis for treatment or patient management decisions.

Negative results should be considered in the context of the individual's recent exposure history, medical history and the presence of clinical signs and symptoms consistent with SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/ Mycoplasma pneumoniae and confirmed by nucleic acid testing as necessary for patient management.

(Invalid)

If any of the control (C) lines do not appear, the test must be interpreted as invalid.

An invalid test result means that your test has encountered an error and the results cannot be interpreted. You will need to retest using a new test card.



10

All used test components should be disposed of in your household waste. After completing all sampling and testing steps, wash hands or use hand sanitizer.



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BioTeke



USER INSTRUCTION

For anterior nasal swabs or oropharyngeal swabs.
SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit

PRODUCT NAME

SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit

PACKAGE SPECIFICATION

1 Test/Kit; 20 Tests/Kit

INTENDED USE

This kit is only used for the in vitro qualitative detection of respiratory multipathogen antigen SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Myoplasma pneumoniae from human oropharyngeal swab specimens.

TEST PRINCIPLE

The kit is immunochromatographic and uses double-antibody sandwich method to detect SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Myoplasma pneumoniae antigen. During detection, the treated specimens are loaded into the sample wells of the test card. When the concentration of SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Myoplasma pneumoniae antigen in specimen is higher than the minimum detection limit, the viral antigen will form complexes with labeled antibodies first. Under chromatography, antigen-antibody complexes move forward along the nitrocellulose membrane till captured by pre-coated monoclonal antibody of SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Myoplasma pneumoniae in the detection zone on the nitrocellulose film to form a red or blue reaction line on the detection zone indicating the test result is positive. Conversely, if there is no viral antigen or the concentration of antigen in specimen is below the minimum detection limit, no red/blue reaction line appears in the detection zone, and the test result is negative. Regardless of whether the sample contains viral antigens or not, a dark blue/purple reaction line will appear in the quality control zone (C). This is the relevant criterion for determining the chromatography process as "normal".

MATERIALS PROVIDED

The test kit consists of test card, sample extraction tube, tube cap, swab and waste bag.

Components	Main Ingredients	Loading quantity (Specification)	
		1 Test/Kit	20 Tests/Kit
Test card	Test strip containing specific SARS-CoV-2/Influenza A virus/Influenza B virus/ Human Parainfluenza virus/Respiratory syncytial virus/Adenovirus/Myoplasma pneumoniae monoclonal antibody, Anti-mouse IgG polyclonal antibody	1pc	20pcs
	Sample extraction tube	1pc	20pcs
Tube cap	Tube cap	1pc	20pcs
	Swab	1pc	20pcs
Waste bag	Waste bag	1pc	20pcs

Note:
1. Test cards are sealed together with desiccant in an aluminum foil pouch.
2. Do not use different batches of test cards and sample extraction tubes.

STORAGE CONDITIONS AND SHELF LIFE

The test card and sample extraction tube should be stored at 2°C-30°C, to be valid for 24 months. Test cards should be used as soon as possible (maximum time: within 1 hour) after opening the foil pouch. The bottle of the sample extraction tube should be capped immediately after use and stored at 2°C-30°C. Only use it within the validity period. Date of manufacture and expiration: See package label for details.

SPECIMEN REQUIREMENTS

The swab specimen should be tested immediately after collection.

LIMITATIONS OF THE TEST

1. The test results of this kit can only serve as reference for clinicians and should not be used as the sole basis for a clinical diagnosis and treatment. Clinical management of patients should be included in the context of their signs and symptoms, their medical history and other laboratory tests, and response to treatment.

2. The quality of the sampling technique and the specimen processing have a greater impact on the detection of pathogens included in this test kit. Thus, a negative test result does not exclude the possibility of a viral infection.
3. Due to methodological limitations of antigen-based test, the analytical sensitivity of immunochromatographic tests is generally lower than that of nucleic acid-based test. Therefore, any test interpretation should pay high attention to negative results and make a comprehensive judgment based on other test results. If clinically necessary, negative results in should be checked by nucleic acid test or virus culture identification.
4. When the test result is positive, it is recommended to apply other methods such as PCR or viral culture for further confirmation if clinically relevant. If necessary or mandated by authorities, please also consult with your local public health office appropriate action.
5. Specifically, false-negative results may occur, if:
(i) Improper sample collection, transport and processing, or low viral titers in the sample.
(ii) samples were taken too early or too late after infection, so that peak viral titers were missed. Multiple samplings at multiple sites in the same patient may help avoid false negative results. efore, multiple sampling at multiple sites in the same patient may avoid false negatives.

PERFORMANCE CHARACTERISTICS

1. The width of the membrane strip of this kit is not less than 2.5 mm, and the liquid migration speed is not less than 10 mm/min.
2. Negative/positive reference coincidence rate
All the positive references are positive for the corresponding pathogens, which is consistent with the known results of the reference. All the negative references are negative for the corresponding pathogen.
3. Repeatability
Repeated testing was conducted for national or enterprise repeatable reference products for 10 times. The test results were consistent with the known results of the reference products and were uniform in color.
4. Analytical specificity
1) Clinical study

SARS-CoV-2				Influenza A			
		QAstat-Dx Respiratory SARS-CoV-2 Panel				QAstat-Dx Respiratory SARS-CoV-2 Panel	
		Positive	Negative			Positive	Negative
SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit		Positive	134	0	134	Positive	112
		Negative	8	766	774	Negative	12
Total		142	766	908	Total	124	764
Statistic		95%CI		Statistic		95%CI	
Sensitivity		94.37%	(89.20%-97.54%)	Sensitivity		90.32%	(83.71%-94.90%)
Specificity		100.00%	(99.52%-100.00%)	Specificity		100.00%	(99.53%-100.00%)
Total coincidence rate		96.12%	(96.27%-96.62%)	Total coincidence rate		96.68%	(97.70%-99.32%)

Influenza B				Human Parainfluenza virus							
		QAstat-Dx Respiratory SARS-CoV-2 Panel				QAstat-Dx Respiratory SARS-CoV-2 Panel					
		Positive	Negative			Positive	Negative				
SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit		Positive	98	0	98	Positive	117	0	117		
		Negative	11	799	810	Negative	11	790	791		
Total		109		799	908	Total		128		790	908
Statistic		Value		95%CI		Statistic		Value		95%CI	
Sensitivity		89.91%		(82.66%-94.85%)		Sensitivity		91.41%		(85.14%-95.63%)	
Specificity		100.00%		(99.54%-100.00%)		Specificity		100.00%		(99.53%-100.00%)	
Total coincidence rate		98.70%		(97.84%-99.39%)		Total coincidence rate		96.70%		(97.84%-99.39%)	

Respiratory syncytial virus				Adenovirus					
		QAstat-Dx Respiratory SARS-CoV-2 Panel	Total			QAstat-Dx Respiratory SARS-CoV-2 Panel	Total		
		Positive	Negative			Positive	Negative		
SARS-CoV-2/Flu A+B/HPIV/RSV/ADV/MP Antigen Rapid Test Kit		Positive	125	0	125	Positive	111	0	111
		Negative	10	773	783	Negative	13	764	797
Total		135	773	908	Total	124	764	908	
Statistic		95%CI		Statistic		Value		95%CI	
Sensitivity		92.59%	(86.80%-96.39%)	Sensitivity		89.52%	(82.74%-94.30%)		
Specificity		100.00%	(99.52%-100.00%)	Specificity		100.00%	(99.53%-100.00%)		
Total coincidence rate		98.95%	(97.98%-99.47%)	Total coincidence rate		98.57%	(97.56%-99.24%)		

M.Pneumonia		QAstat-Dx Respiratory SARS-CoV-2 Panel		Total
		Positive	Negative	
SARS-CoV-2/Flu A+B nPIR/RSV/ADV/MP Antigen Rapid Test Kit	Positive	105	0	105
	Negative	9	794	803
Total		114	794	908

2) Cross-reactivity
There is no cross-reactivity with the following pathogens:

No.	Virus/ Bacteria name	Strain	Concentration / CT value
1	Coronavirus HKU I	GUI804-138	CT: 23

No.	Virus/ Bacteria name	Strain	Concentration / CT value
2	Coronavirus OC43	VR-1558, OC43	4.2×10 ⁵ TCID ₅₀ /mL
3	Coronavirus NL63	NL63	1.6×10 ³ TCID ₅₀ /mL
4	Coronavirus 229E	229E/GZ/1801-3	5.6×10 ⁴ TCID ₅₀ /mL
5	Rhinovirus (group A)	A30/GZ/1710-89	4.2×10 ⁶ TCID ₅₀ /mL
6	Rhinovirus (group B)	70/F02-2547	1.0×10 ⁶ TCID ₅₀ /mL
7	Enterovirus (CA16)	CA16 /Guangzhou/0302/2011	1.8×10 ⁷ TCID ₅₀ /mL
8	Enterovirus (Echo)	ATCC VR-39, HILL	1.0×10 ⁶ TCID ₅₀ /mL
9	Enterovirus (EV71)	EV71/Guangzhou/0402/012	5.6×10 ⁴ TCID ₅₀ /mL
10	Epstein-barr virus capsid antigen	B95-8	CT: 17
11	Measles virus	Edmonston	1.0×10 ⁷ TCID ₅₀ /mL
12	Human cytomegalovirus	RC256	3.2×10 ³ TCID ₅₀ /mL
13	Rotavirus	VR-2018	CT: 20
14	Norovirus	ATCC VR-3234SD	3.6×10 ⁶ Copies/mL
15	Mumps virus	Jones	1.0×10 ⁷ TCID ₅₀ /mL
16	Varicella zoster virus	VR-1367	CT: 13
17	MERS-coronavirus	EMC/2012	1.6×10 ⁵ TCID ₅₀ /mL
18	Human metapneumovirus	GZ/1803-107	1.0×10 ⁵ TCID ₅₀ /mL
19	Haemophilus influenzae	GIM 1.961.	4.8×10 ⁷ CFU/mL
20	Chlamydia pneumoniae	ATCC VRJ-2282, TW183	4.2×10 ⁶ TCID ₅₀ /mL
21	Streptococcus pyogenes	ATCC 19615	1.6×10 ⁸ CFU/mL
22	Pooled human pharyngeal washes	N/A	100%
23	Bordetella pertussis	GDM 1.952	2.6×10 ⁸ CFU/mL
24	Legionella pneumophila	Philadelphial, Brenner	1.9×10 ⁶ CFU/mL
25	Staphylococcus aureus	CMCC(B) 26003	2.6×10 ⁸ CFU/mL
26	Staphylococcus epidermidis	191 (Winslow and Winslow) Evans	7.7x10 ⁵ CFU/mL
27	Candida albicans	CMCC(F) 129002	1.3x10 ⁵ CFU/mL
28	Streptococcus pneumoniae	(Klein) Chester	1.0×10 ⁶ CFU/mL

BioTeke test detects all the pathogens listed below: SARS-CoV-2, Influenza A virus, Influenza B virus, Respiratory syncytial virus, Adenovirus and Mycoplasma pneumoniae.

No.	Virus/Bacteria name	Strain	Concentration/ CT value
1	Influenza A virus 2009H1N1	L19-A1/Si chuan/SWL1/2009	4.2×10 ⁶ TCID ₅₀ /mL
2	Influenza A virus seasonal H1N1	L6-A1/ Liaoning/huanggu /1183/2007	5.6×10 ⁵ TCID ₅₀ /mL
3	Influenza A virus H3N2	L8-A3/ Brisbane/10/2007	1.0×10 ⁶ TCID ₅₀ /mL
4	Influenza A virus H5N1	A/Chicken/Liaoning/SD007/ 2017(H5N1)	CT: 20
5	Influenza A virus H7N9	A/Guangd/17SF003/2016(H7 N9)	CT: 20

6	Influenza B virus Yamagata	GZ/174/201803	5.6×10 ⁶ TCID ₅₀ /mL
7	Influenza B virus Victoria	GZ/133/201712	1.0×10 ⁶ TCID ₅₀ /mL
8	Respiratory syncytial virus A	RSVA/GZ/Hecin170574	1.3×10 ⁷ TCID ₅₀ /mL
9	Respiratory adenovirus type I	ADVIGZ/Hecin160821	2.4×10 ⁷ TCID ₅₀ /mL
10	Respiratory adenovirus type 2	GUI705-34/2017	5.6×10 ⁷ TCID ₅₀ /mL
11	Respiratory adenovirus type 3	ADV3/GZ/0101/2011	1.0×10 ⁸ TCID ₅₀ /mL
12	Respiratory adenovirus type 4	ADV4/GZ/Hecin161172/2016	5.6×10 ⁷ TCID ₅₀ /mL
13	Respiratory adenovirus type 5	ADV/GZ/1801-54	1.0×10 ⁷ TCID ₅₀ /mL
14	Respiratory adenovirus type 7	ADV7/GZ/1706-198	3.2×10 ⁷ TCID ₅₀ /mL
15	Respiratory adenovirus type 55	ADV55/GZ/1612-129	3.2×10 ⁷ TCID ₅₀ /mL
16	SARS-CoV-2	Wild Type	2.8×10 ⁶ TCID ₅₀ /mL
17	Mycoplasma pneumoniae	ATCC 15531	1.0×10 ⁹ Copies/mL
18	Human Parainfluenza virus 1	PIV1/Guangzhou/07011	1.3×10 ⁷ TCID ₅₀ /mL
19	Human Parainfluenza virus 2	PIV2/GZ/Hecin171134/2017	5.6×10 ⁷ TCID ₅₀ /mL
20	Human Parainfluenza virus 3	PIV3/Guangzhou/0903/2012	3.2×10 ⁷ TCID ₅₀ /mL
21	Human Parainfluenza virus 4a	ATCC VR-1378, M-25	4.5×10 ⁷ TCID ₅₀ /mL
22	Human Parainfluenza virus 4b	ATCC VR-1377, CH19053	1.3×10 ⁷ TCID ₅₀ /mL